学位論文の内容の要旨

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Summary	or tn	e Substanc	e of Di	ssertation

専 攻 Major Field	分子情報制御学	部 門 Department	分子 腫瘍学 。
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Summary

Dental caries are an important global health concern, and Streptococcus mutans has been established as a major cariogenic bacterial species. Reports indicate that a rare sugar, D-tagatose, is not easily catabolized by pathogenic bacteria. In this study, we examined the inhibitory effects of D-tagatose on the growth and biofilm formation of S. mutans GS-5. Monitoring the S. mutans growth over a 24-h period showed that D-tagatose prolonged the lag phase without interfering with the final cell yield. This growth retardation was also observed in the presence of 1% sucrose, although it was abolished by the addition of D-fructose. S. mutans biofilm formation was significantly inhibited by D-tagatose (1.0 to 4.0%) compared with that in the culture containing sucrose alone, and S. mutans formed granular biofilms in the presence of this rare sugar. The inhibitory effect of D-tagatose on S. mutans biofilm formation was more evident than that of xylitol. The addition of 1% D-tagatose significantly decreased the expression of gtfB, fruA, and D-fructose-specific phosphotransferase genes but not the expression of ftf compared with the culture containing 1% sucrose. The activity of cell-associated glucosyltransferase in S. mutans was inhibited by 4% D-tagatose. These results indicate that D-tagatose reduces water insoluble glucan production from sucrose by inhibiting glucosyltransferase activities, which limits access to the free D-fructose released during this process and retards the growth of S. mutans.

Materials and Methods

S.mutans GS-5 was incubated in presence of D-tagatose for checking growth and pH. Biofilm was formed and stained with crystal violet for assay. Moreover, biofilm was formed over a plastic disc to study under scanning electron microscope (SEM). Real time PCR analysis was performed to observe sugar metabolism gene expression in S.mutans GS-5 during incubation with D-tagatose. Cell associated GTF B enzyme was extracted and incubated with D-tagatose to check the glucan production.

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Results; Effects of D-tagatose on S. mutans GS-5 growth

First, the effects of D-tagatose on S. mutans GS-5 growth in BHI containing 1% sucrose were examined. Sucrose enhanced S. mutans GS-5 growth compared with BHI alone, and the pH of the culture decreased to less than 5.0 after 9-h incubation. Interestingly, D-tagatose delayed the transition of S. mutans growth to the logarithmic phase despite the presence of sucrose. Correspondingly, the pH decline of the culture was also delayed by

D-tagatose compared with that in sucrose alone.

Effects of D-tagatose on in vitro S. mutans GS-5 biofilm formation

We evaluated the effects of D-glucose, xylitol, and D-tagatose on in vitro S. mutans GS-5 biofilm formation. The addition of 1% sucrose markedly promoted biofilm formation by S. mutans GS-5, which is consistent with many previous reports. Xylitol, D-tagatose, and their combination significantly reduced S. mutans GS-5 biofilm formation compared with 1% sucrose alone or when supplemented with 1% glucose.

Scanning electron microscopy examination of S. mutans GS-5 biofilms

S. mutans GS-5 was cultured in 1 ml of BHI containing 1% sucrose with or without 1.0% or 4.0% each of xylitol or D-tagatose in 24-well plates with plastic disc inserts. The plates were incubated anaerobically at 37°C for 72 h, and the biofilms formed on the plastic discs were compared. Interestingly, many S. mutans GS-5 cell aggregates were observed in the culture containing D-tagatose (especially 4.0%), whereas homogeneous biofilms formed on the discs in the other 1% sucrose-containing cultures tested.

Effects of D-tagatose on the expression of sucrose metabolism genes in S. mutans GS-5

We also confirmed that 1% sucrose increased the expression of gtfB but did not increase the expression of the fructosyltransferase gene (ftf) compared with bacteria cultured in BHI alone. By contrast, the induction of gtfB expression by sucrose was strongly inhibited by D-tagatose (1%). The expression of the exo-B-fructosidase gene (fruA), which releases free D-fructose from FTF-producing water-soluble fructan, was also reduced in the presence of D-tagatose. Among the phosphotransferase system (PTS) genes involved in metabolizing sucrose or sucrose-derived monosaccharide (D-glucose and D-fructose), the expression levels of ptsfru, ptsfru/man and ptsglu/man were increased by 1% sucrose, although this increase was inhibited by D-tagatose. GtfB releases D-fructose during glucan synthesis, and these insoluble glucans are degraded by dextranase and utilized as a glucose source. The fruA gene product releases free D fructose from the fructan synthesized by FTF. The repression of these genes as well as the fructose or glucose pts genes indicates that D-tagatose limits the ability of S. mutans GS-5 to access sucrose-derived monosaccharides, especially D-fructose. Shows that D-fructose is a powerful inducer of gtfB expression, and sucrose strongly increases the expression of fructose PTS, thus indicating that D-fructose is a key sugar required for S. mutans growth and biofilm formation.

Effects of D-tagatose on S. mutans GS-5 GTF activity

S. mutans GTFs B and C, which produce water insoluble glucan, are known to be cell-associated. To examine whether D-tagatose directly inhibits the activity of GTF in S. mutans GS-5, the cell-associated proteins were extracted with urea from S. mutans GS-5 cells cultured in BHI. The water insoluble glucan production was compared among the renatured enzymes in media with 0.1 M sucrose in the presence or absence of 4% D-tagatose. The addition of 4.0% D-tagatose significantly decreased the water insoluble glucan production, indicating that the sugar directly inhibits the activity of S. mutans GS-5 cell-associated GTF.

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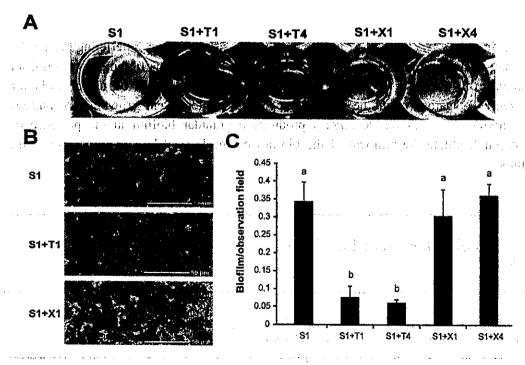
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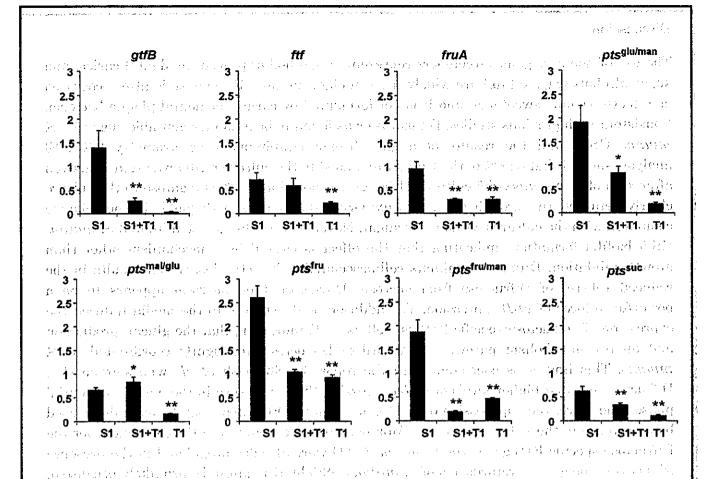
Disscussion

The use of non-cariogenic sweeteners represents a method of preventing dental caries, and sugar alcohols (e.g., xylitol) are widely used in chewing gum. D-tagatose is also recognized as a tooth-friendly sweetener, and it is not fermented by cariogenic dental plaque bacteria. Consistent with previous studies, D-tagatose was found to be a non-fermentable sugar for S. mutans GS-5, and the results of a gas chromatography-mass spectrometry (GC-MS) analysis showed that 81.6% of the D-tagatose added to the culture media was retained, even after 48 h of S. mutans GS-5 culture. Although the addition of 1% D-tagatose to the culture clearly retarded the growth of S. mutans GS-5, the final growth yield did not change compared with the cultures without the sugar. Nevertheless, D-tagatose inhibited S. mutans GS-5 biofilm formation, indicating that the effect is caused by a mechanism other than growth inhibition. D-tagatose inhibits cell-associated GTF activities, which results in the reduced release of D-fructose from sucrose. D-fructose (and sucrose) appears to be a powerful inducer of gtfB expression. The addition of 1% sucrose to the media induced the expression of D-fructose-specific PTS as well as gtfB, indicating that the glucan production and energy metabolism pathways that utilize D-fructose are tightly coordinated in S. mutans. This finding is consistent with the report by Shemesh et al., who showed that D-fructose induced higher levels of gtfB expression than D-glucose in the early exponential phase. Therefore, the suppression of gtfB expression by D tagatose may be partially caused by a decrease in the D-fructose supply. Moreover, genes encoding the EII component for the D-fructose-specific PTS genes (ptstru and ptstru/man) were also downregulated in the presence of sucrose. The growth retardation of S. mutans GS-5 by D-tagatose is also likely because of the limited D fructose supply resulting from GTF inhibition because the D tagatose induced growth retardation was reversed under D-fructose supplementation. We predict that changes in the availability of this monosaccharide are responsible for the prolongation of the lag-phase S. mutans GS-5 growth by D-tagatose.

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Figure; S.mutans biofilm inhibition by D-tagatose. Here, S1- 1% Sucrose, S1+T1- 1% Sucrose+ 1% D-tagatose, S1+T4- 1% Sucrose+ 4% D-tagatose, S1+X1- 1% Sucrose+ 1% Xylitol, S1+X4- 1%Sucrose+ 4% Xylitol. Clear biofilm inhibition has seen by D-tagatose



Figure; Effect of D-tagatose on S. mutans sugar metabolism gene expression. Here, S1-1% Sucrose, S1+T1-1% Sucrose+1% D-tagatose, T1-1% D-tagatose. The expression was measured in BHI media.

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Conclusion

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D-tagatose appears to inhibit S. mutans GS-5 growth and biofilm formation by interfering with GTF activity. This effect may be useful in the prevention of dental caries. Based on the findings obtained from this study, we conclude that foods or preparations containing D-tagatose could be useful tools for improving oral hygiene. D-tagatose might be able to suppress the intermittent growth of S. mutans between oral care activities. In addition, S. mutans produces a granular biofilm in the presence of D-tagatose, which might facilitate the removal of the biofilm by mechanical brushing compared with homogeneous biofilms.

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